



Are ranaviruses host specific or not?



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Introduction

Ranaviruses are a group of systemic iridoviruses associated with systemic disease in mainly erythropoietic tissue, potentially causing high mortalities in affected animals. Ranaviruses have been isolated from fish, amphibians and reptiles in Austral-Asia, North America and Europe. Even though ranaviruses are often named according to the host from which they were originally isolated, little is known about the host specificity of these viruses

Methodology

The aim of this study was to identify which species of fish are susceptible to infection with *Ranavirus* and whether ranaviruses are host specific or not. This was investigated by a literature search of initial isolations of ranaviruses from fish and of experimental challenges of susceptibility of fish to ranaviruses.

For the challenge experiments, species were either designed as susceptible (Y), Infectable (I) or Non-susceptible (N) according to the following definitions: Y1: Statistically significant mortalities, Y2: High mortality and pos. virus re-isolation, Y3: High mortality, Y4: Mortality >15% and pos. virus re-isolation, I1: Mortality <5% and pos. virus re-isolation, I2: No statistically significant mortality but pos. virus re-isolation, N1: No statistically significant mortality and no virus re-isolation, N2: No mortality and no virus re-isolation, N3: Mortality <10% (For Y3 and N3 there was no data on virus re-isolation).

The results of the study can be seen in tables I & II.

Table I. Published findings of natural outbreaks of ranavirus in fish (Only *Ranavirus* isolates confirmed by sequencing are included)

| Fish species | Isolate | <i>Ranavirus</i> species* | Location(s) | Origin of fish | Mortality | Age of fish | Year of outbreak(s) | Reference(s) |
|--|-------------------------------------|---------------------------|-------------------------|----------------|-----------|-----------------|---------------------|--------------|
| Redfin perch (<i>Perca fluviatilis</i>) | Epizootic haematopoietic EHN | EHN | Australia | Wild | High | Mostly juvenile | 1984 -> 1994 | 11, 20 |
| Rainbow trout (<i>Oncorhynchus mykiss</i>) | Epizootic haematopoietic EHN | EHN | Australia | Farmed | 0-0.1% | All ages | 1986, 1993 & 1996 | 12, 21 |
| Sheatfish (<i>Silurus glanis</i>) | European sheatfish virus (ESV) | ECV | Germany | Farmed | 30-100% | Fry and adults | 1988, 1990 | 1, 2 |
| Catfish (<i>Ictalurus melas</i>) | European catfish virus | ECV | France, Italy | Farmed | 90-100% | All ages | 1990, 1993 | 6, 17 |
| Doctor fish (<i>Labroides dimidiatus</i>) | Doctor fish virus (DFV) | SCRV | Imported from Singapore | Farmed | ND | ND | 1995 | 9 |
| Guppy (<i>Poecilia reticulata</i>) | Guppy virus 6 (GV6) | SCRV | Imported from Singapore | Farmed | ND | ND | 1995 | 9 |
| Tilapia (<i>Oreochromis mossambicus</i>) | Bohle iridovirus** | BIV | Australia | Farmed | 100% | Fry | 1996 | 3 |
| Threespine stickleback (<i>Gasterosteus aculeatus</i>) | Redwood creek virus (RCV)** | FV3 | California | Wild | 20% | ND | 1996 | 14 |
| Largemouth bass (<i>Micropterus salmoides</i>) | Largemouth bass iridovirus (LMBV) | SCRV | USA | Wild | ND | Adult | 1996 | 15 |
| Pike-perch (<i>Sander lucioperca</i>) | Pike-perch iridovirus (PPV) | Unresolved | Finland | Farmed | 0% | Fingerlings | 1998 | 19 |
| Grouper (<i>Epinephelus spp.</i>) | Singapore grouper iridovirus (SGIV) | Tentative | Singapore | Farmed | >90% | Fry and adults | 1998 | 8, 18 |
| Shortfinned eel (<i>Anguilla australis</i>) | Shortfinned eel ranavirus (SERV) | Unresolved | Imported to Italy | Farmed | 0% | Juvenile | 1999 | 7 |

*According to International Committee on Taxonomy of viruses. EHN=epizootic haematopoietic necrosis virus, ECV=european catfish virus, SCRV=Santee-cooper ranavirus, BIV=Bohle iridovirus, FV3= Frog virus 3.
**Amphibian ranaviruses
ND= No data available

Discussion

In half of the initial ranavirus-isolations, no mortalities were observed. In two cases were amphibian ranaviruses found in association with mortalities in fish, and EHNV has caused natural outbreaks in two fish species, although with different mortalities. EHNV is the most investigated *Ranavirus*, and can infect 12 different fish species, even though some only by injection.

The study showed that most of the challenge studies has been performed with very low numbers of fish, and often no information about virus re-isolation is given. This makes it difficult to determine the susceptibility of a fish species to a specific *Ranavirus*.

Table II. Published challenge trials with ranaviruses in fish. Challenge trials with fish species already listed in table I are not included, unless it has been challenged with another ranavirus than the one it was naturally infected with.

| Fish species | Virus isolate | Location | Age of animals | Type of challenge* | Temperature in challenge (°C) | No. of animals challenged | Mortality (%) | Morbidity (%) | Percent positive virus re-isolations | Reference | Susceptibility** |
|--|---------------|------------|----------------|--------------------|-------------------------------|---------------------------|---------------|---------------|--------------------------------------|-----------|------------------|
| Atlantic salmon <i>Salmo salar</i> | EHN | Australia | Juvenile | IP | NS | 15 | 0 | 100% | NS | 11 | Y3 |
| Barramundi <i>Lates calcarifer</i> | EHN | Australia | 4 mths | Bath | 18-20 | 10 | 0 | 0 | NS | 13 | N3 |
| | | | | IP | 18-20 | 3 | 0 | 0 | | | N3 |
| | BIV | Australia | NS | IP+IM | 18-20 | 20 | 100 | 0 | 0 | 4 | Y3 |
| | BIV | Australia | Juvenile | IP+IM | 29 | 20 | 100 | 0 | 75-100 | 16 | Y2 |
| | | | | Bath | 29 | 20 | 100 | 0 | 50-100 | | Y2 |
| Channel catfish <i>Ictalurus punctatus</i> | EHN | California | Fry | Bath | 15 | 20 | 0 | 0 | 0 | 9 | N2 |
| | ESV | | | Bath | 15 | 20 | 0 | 0 | 0 | | N2 |
| | DFV/GV | | | Bath | 15 | 20 | 0 | 0 | 0 | | N2 |
| Chinook salmon <i>Oncorhynchus tshawytscha</i> | EHN | | Fry | Bath | 15 | 17 | 18 | 0 | NS | | N3 |
| | ESV | | Fry | Bath | 15 | 17 | 0 | 0 | 0 | | Y4 |
| | DFV/GV | | Fry | Bath | 15 | 17 | 6 | 0 | 0 | | N3 |
| Rainbow trout <i>Oncorhynchus mykiss</i> | PPV | Finland | >1 year | IP | 15-19 | 80 | 0 | 0 | 43 | 19 | I1 |
| | | | | Bath | 15-19 | 40 | 0 | 0 | 0 | | N2 |
| | ESV | California | Fry | Bath | 15 | 25 | 4 | 0 | NS | 9 | N3 |
| | DFV/GV | | | Bath | 15 | 25 | 4 | 0 | 0 | 10 | N3 |
| | ATV | USA | Adults | Oral | NS | 12 | 0 | 0 | 0 | 10 | N2 |
| | BIV | Australia | | IP+IM | NS | 0 | 0 | 0 | NS | 4 | N3 |
| Tilapia <i>Oreochromis mossambicus</i> | BIV | Australia | Fry | Oral | 24 | 12 | 30+ | 0 | 0 | 3 | Y4 |
| | EHN | Australia | 6 mths | Bath | 18-20 | 5 | 0 | 0 | 0 | 13 | N2 |
| | | | | IP | 18-20 | 5 | 20 | 20 | frommorts | | Y4 |
| Australian bass <i>Macquaria novemcincta</i> | EHN | Australia | 2 mths | Bath | 18-20 | 10 | 0 | 0 | 0 | 13 | N2 |
| | | | | IP | 18-20 | 6 | 100 | 100 | 100 | | Y2 |
| Golden perch <i>Macquaria ambigua</i> | EHN | Australia | 3 mths | Bath | 18-20 | 10 | 100 | 100 | 100 | 13 | Y2 |
| | | | | IP | 18-20 | 5 | 100 | 100 | 100 | | Y2 |
| Macquarie perch <i>Macquaria australasica</i> | EHN | Australia | 2 mths | Bath | 18-20 | 4 | 0 | 0 | 50 | 13 | I1 |
| | | | | IP | 18-20 | 4 | 100 | 100 | 100 | | Y2 |
| Murray cod <i>Maccullochella peelii</i> | EHN | Australia | 3 mths | Bath | 18-20 | 10,10,6 | 30-66 | 30-66 | frommorts | 13 | Y4 |
| | | | | IP | 18-20 | 5 | 100 | 100 | 100 | | Y2 |
| Silver perch <i>Bidyanus bidyanus</i> | EHN | Australia | >1 year | Cohab | 18-20 | 10 | 0 | 0 | 0 | 13 | N2 |
| | | | | Bath | 18-20 | 12 | 0 | 0 | 0 | | N2 |
| Smelt <i>Retropinna semoni</i> | ATV | USA | Adults | IP | 8 | 0 | 0 | 0 | 0 | 10 | N2 |
| | | | | Oral | 15 | 20 | 0 | 0 | 0 | | Y3 |
| Sunfish <i>Lepomis cyanellus</i> | EHN | Australia | <1 year | Bath | 18-20 | 8 | 0 | 0 | 0 | 13 | N2 |
| | | | | IP | 18-20 | 3 | 0 | 0 | 0 | | N2 |
| Goldfish <i>Carassius auratus</i> | EHN | Australia | >1 year | Cohab | 18-20 | 4 | 100 | 100 | 100 | 13 | Y2 |
| | | | | Bath | 18-20 | 10 | 90 | 90 | frommorts | | Y2 |
| Mosquito fish <i>Gambusia affinis</i> | ATV | USA | Adults | IP | 16 | 0 | 0 | 0 | 0 | 10 | N2 |
| | | | | Oral | 25 | 4 | 0 | 0 | 0 | | N2 |
| Tiger barb <i>Capoeta terezonca</i> | EHN | Australia | <1 year | Bath | 18-20 | 5 | 0 | 0 | 0 | 13 | N2 |
| Pike <i>Esox lucius</i> | EHN | Denmark | Fry | Bath | 12 | 96 | 100 | 0 | 95 | 5 | Y1 |
| | | | | | 22 | 94 | 98 | 0 | 82 | | I2 |
| | ESV | | | | 12 | 107 | 73 | 0 | 54 | | Y1 |
| | | | | | 22 | 185 | 97 | 0 | 94 | | I2 |
| | ECV | | | | 12 | 98 | 51 | 0 | 37 | | I2 |
| | | | | | 22 | 99 | 98 | 0 | 68 | | I2 |
| | PPV | | | | 12 | 83 | 61 | 0 | 84 | | Y1 |
| | | | | | 22 | 101 | 93 | 0 | 100 | | I2 |
| | SERV | | | | 12 | 102 | 100 | 0 | 82 | | Y1 |
| | | | | | 22 | 70 | 97 | 0 | 0 | | I2 |
| | | | | | 12 | 81 | 44 | 0 | 60 | | N1 |
| | FV3 | | | | 22 | 69 | 83 | 0 | 60 | | I2 |
| Mountain galaxias <i>Galaxias olidus</i> | EHN | Australia | >1 year | Bath | 18-20 | 5 | 100 | 100 | 100 | 13 | Y2 |

*IP=intraperitoneal injection, IM=intramuscular injection

** See explanation in text
NS=not stated

References

- Ahne et al 1989 J Vet Med B, 36, 333-36
- Ahne et al 1991 Bull EAEP, 11, 3, 97-8
- Ariel & Owens 1997 Dis Aquat Org, 29, 1, 1-6
- Ariel et al 1995 Diseases in asian aquaculture II, 355-67
- Bang Jensen et al 2009 Dis Aquat Org, 83, 3, 169-79
- Bovo et al. 1993 Boll Soc It Patol Ittica, 11, 3-10
- Bovo et al 1999 9th Int Con EAEP, P-153
- Gibson-Kueh et al 2003 J Comp Path, 129, 111-19
- Hedrick & McDowell 1995 Vet Res, 26, 423-27
- Jancovich et al 2001 Dis Aquat Org, 46, 159-63
- Langdon et al 1986 J Fish Dis, 9, 263-68
- Langdon et al 1988 J Fish Dis, 11, 93-6
- Langdon 1989 J Fish Dis, 12, 295-310
- Mao et al 1999 Vir Res, 63, 1-2, 45-52
- Mao et al 1999 Dis Aquat Org, 37, 107-14
- Moody & Owens 1994 Dis Aquat Org, 18, 95-102
- Pozet et al 1992 Dis Aquat Org, 14, 35-42
- Qin et al 2003 Dis Aquat Org, 53, 1-9
- Tapiovaara et al 1998 Dis Aquat Org, 32, 185-193
- Whittington et al 1996 Aus Vet J 73, 112-14
- Whittington et al 1999 Dis Aquat Org, 35, 125-130