ASSESSMENT OF BIOAEROSOLS IN SWINE BARNS







Correia-Gomes, C.^{1, 3}, Niza-Ribeiro, J.^{1,2,3}



¹ICBAS -Instituto de Ciências Biomédicas Abel Salazar – Universidade do Porto, Largo Prof. Abel Salazar, 2, 4099-003 Porto, PORTUGAL
² Segalab – Laboratório de Sanidade Animal e Segurança Alimentar, Rua de Recarei, Gondivai, 4465-734 Leça do Balio, PORTUGAL
³ ISPUP – Instituto de Saúde Pública da Universidade Porto, Rua das Taipas, 4099-003 Porto, PORTUGAL

BACKGROUND & OBJECTIVE

Poor air quality in livestock buildings adversely affects animal health through a variety of mechanisms, and also enables the transmission of diseases, like Salmonelosis. In bioaerosols we have bacteria and fungi. The Gram negative bacteria (G-) are some of the most important pathogens for swine, and included fecal bacteria, like *Escherichia coli*, *Salmonella* sp., *Neisseria* sp., *Pseudomonas* sp., among others. Some studies have reported G- bacterial exposures in swine barns between 7×10³ and 65×10⁵ CFU/m³ (1), while others (2) reported significantly lower concentrations of 0.42 to 4.52×10² CFU/m³. Salmonella airborne transmission was proved possible by experimental studies (3,5). The infection dose found were higher than 10⁶ CFU and depends of serotype.

AIM: To assess the bioaerosol contamination in swine barns and with this to obtain field information providing an insight into the possible risk of airborne transmission of Salmonella in swine herds. For this purpose **Gram negative bacteria (G-)** were used as an **indicator** of possible Salmonella presence.

MATERIALS & METHODS

DATA:

One herd with post-weaning barns (active ventilation) and two types of fattening barns (passive versus active ventilation).

Samples taken at the passages in the middle of the barn at a height near the nostril of the pigs.

An air volume of 5 liters was aspirated to total bacteria counts and fungi and 30liters to G- counts.

- Samples taken once a month in January, February and April 2009 (winter to spring days).
- Temperature and humidity at the barn and outside the barn was recorded using a hygrometer.
- Potential differences between barns and months were tested.

MICROBIOLOGICAL ANALYSIS:

The air was aspirated by "air IDEAL microbiological air sampler" (bioMérieux®) to 90mm diameter Petri dishes.

Three types of culture media were used: Agar MacConkey for G-, Agar Gelose Tripcase Soja for total bacterial and Agar Sabouraud glucose cloranfenicol for fungi.

✤ The plates were incubated at 30°C/72h to total bacteria, at 37°C/48h to G- and at 25°C/5days (120h) for fungi.

✤ CFUs counts were corrected to most probable number of bacteria (MPN) and divided by the volume of air sampled and are presented as CFU/m³.

RESULTS & DISCUSSION

• Mean stocking densities were 20 pigs/pen in fattening pens (pens with 4x4m) and 25 pigs/pen in post-weaning pens (pens with 2x2m).

• Figure 1 and 2 shows the distribution of CFU/m³ fungi, total bacteria and G- for naturally and mechanically ventilated fattening barn and post-weaning barn.

There were no statistical difference between barns for fungi, total bacteria and G- counts.

Comparing the values of CFU/m³ of fungi along the months tested showed statistical differences (p<0.04) between January and the rest of the months (Figure 3).



✓ High levels of total bacteria and fungi concentration in all barns, similar to the ones found in others studies (4).

Y This indicate a highly contaminated air with could favor the transmission of microbiological agents within the farms. The piglets are especially at risk because they have a poor immunity system and the results in the post-weaning barn were worst than in the others barns.

✓ The contamination with G- was lower than expected. It is lower than the one found in other studies (1) but agrees to with Chang's data (2).

✓ Using the G- bacteria counts as indicator of fecal contamination and of the possible presence of Salmonella, the concentration of Salmonella (if present) will be in lower than G- counts and under the infective dose of Salmonella trough the air, therefore the risk of airborne infection with Salmonella seemed negligible.

Although the number of samples from the study was low we decided not to continue with this study because we felt that the results would not become different then the ones obtained.

REFERENCES

(1) Attwood P, Brouwer R, Ruigewaard P, Versloot P, de Wit R, Heederik D, Boleij JS. 1987. A study of the relationship between airborne contaminants and environmental factors in Dutch swine confinement buildings. Am Ind Hyg Assoc J 48: 745-751.

(2) Chang CW, Chung H, Huang CF, Su HJ. 2001. Exposure of workers to airborne microorganisms in open-air swine houses. Appl Environ Microbiol 67: 155-161.

(3) Oliveira CJ, Carvalho LF, Garcia TB. Experimental airborne transmission of Salmonella Agona and Salmonella Typhimurium in weaned pigs. Epidemiology and infection 2006; 134(1): 199-209.

(4) Predicala BZ, Urban JE, Maghirang RG, Jerez SB, Goodband RD. 2002. Assessment of bioaerosols in swine barns by filtration and impaction. Curr Microbiol 44: 136-140. (5) Proux K, Cariolet R, Fravalo P, Houdayer C, Keranflech A, Madec F. Contamination of pigs by nose-to-nose contact or airborne transmission of Salmonella Typhimurium. Veterinary research 2001; 32(6): 591-600.

Acknowlegments:

SVEPM's contribution to the delegate conference attendance bioMérieux[®] Farmer and veterinarian that participated in this study